Vol. 106, No. 2, 1982 BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS
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ERRATA

Volume 105, Number 1, March 15, 1982

In "The Rapid PI-Turnover Is Not Coupled with the Aggregation in A23187-Activated Human Platelets," by Atsushi Imai and Yoshinori Nozawa, pages 236-243, in Figures 2-4, on pages 239-241, respectively, the units of incubation time were written incorrectly. The units on the abscissas should read "INCUBATION TIME (sec)" instead of "INCUBATION TIME (min)."

Volume 104, Number 4, February 26, 1982

In "Integration of Purified Adrenocortical Cytochrome P-$450_{11\beta}$ into Phospholipid Vesicles," by A. Lombardo, G. Defaye, C. Guidicelli, N. Monnier, and E. M. Chambaz, pages 1638-1645, on page 1642, Figure 3 was incorrectly reproduced. A proper reproduction appears on the following page for the reader's convenience.

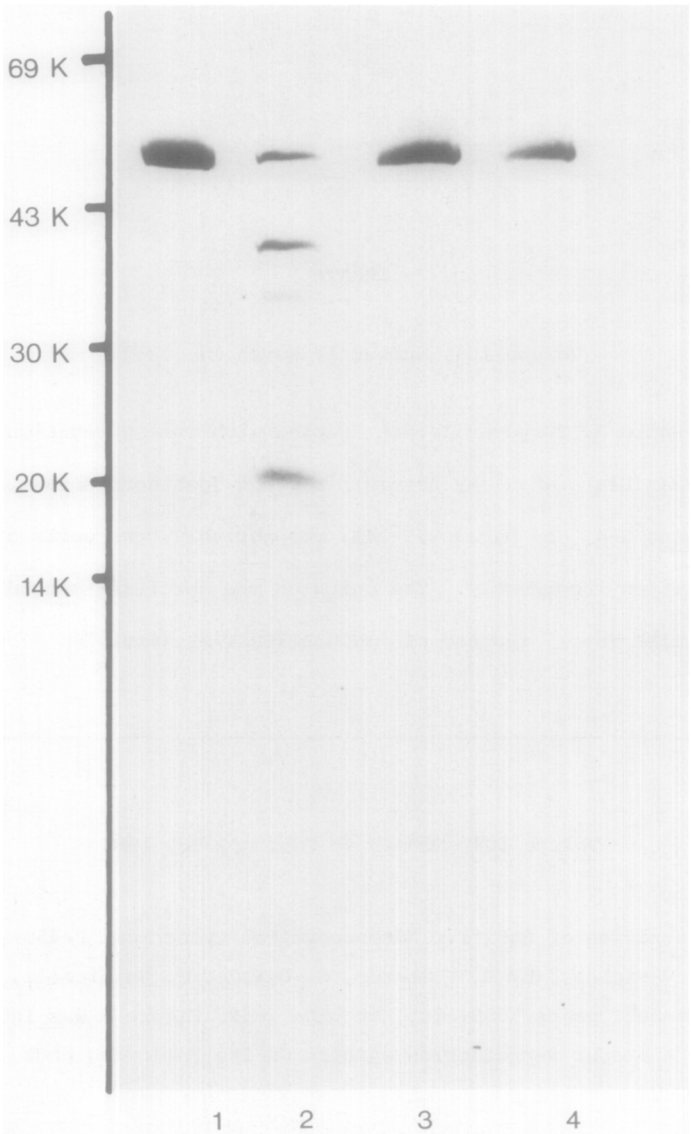

Fig. 3 - SDS (0.1%), polyacrylamide (15%) gel electrophoresis of various cytochrome P-450$_{11\beta}$ preparations. Lane 1: purified cyt P-450$_{11\beta}$ after octyl Sepharose chromatography; lane 2: purified cyt P-450$_{11\beta}$ after treatment with trypsin; lane 3: liposome-incorporated cyt P-450$_{11\beta}$ as obtained after Sepharose 4B filtration (see fig. 2); lane 4: same preparation as in lane 3, after treatment with trypsin as for cyt P-450$_{11\beta}$. Lanes 2 and 4: trypsin was used at 0.2% (v/w) for 2.5 h at 20°C.

Volume 105, Number 2, March 30, 1982

In "Evidence for Two Classes of Rat Plasma Fibrinogen γ Chains Differing by Their COOH-Terminal Amino Acid Sequences," by C. D. Legrele, C. Wolfenstein-Todel, Y. Hurbourg, and M. W. Mosesson, pages 521-529, Figures 1-4, on pages 523-526, respectively, appeared with incomplete legends. For the reader's convenience, the figures and complete legends appear below.

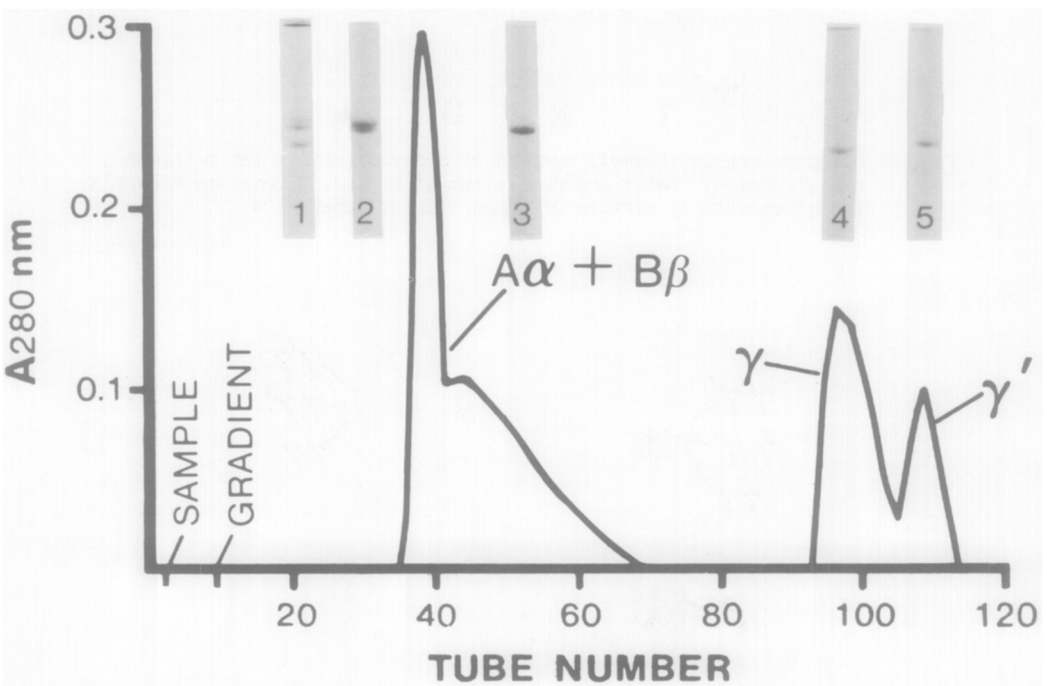

Fig. 1 DEAE-cellulose chromato-graphy of 22 mg S-carboxymethyl rat fibrinogen using a gradient (9 chambers, 75 ml each) from 0.01M to 0.2M tris-phosphate in 8M urea at pH 7.0. The times of sample and gradient application are indicated; fractions of 6 ± 0.2 ml were collected. The types of chains contained within each peak are indicated. Weber and Osborn gels are shown: starting material, 1; first peak, 2; second peak, 3; third peak (γ), 4; fourth peak (γ'), 5.

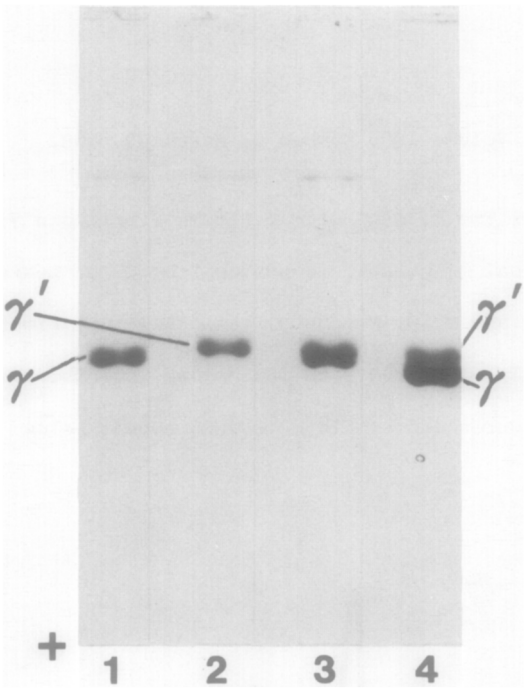

Fig. 2 Electrophoresis (Laemmli system) of S-carboxymethyl rat or human γ and γ' chains. Rat γ chains, 1; rat γ' chains, 2; mixture of rat γ and γ' chains, 3; mixture of human γ and γ' chains, 4.

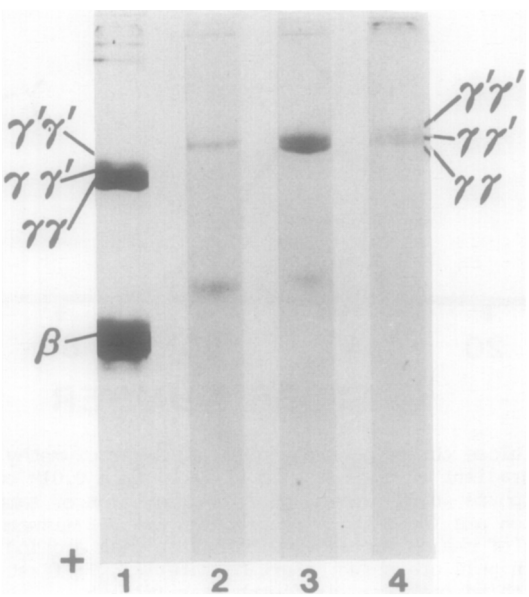

Fig. 3 Electrophoresis (Laemmli system) of reduced, crosslinked rat fibrin (gel 1) and S-carboxylmethylated material (separate analysis) from the γ-chain peak of the column shown in Fig. 4 (γ chain peak, ascending limb, 1; same, middle of peak, 3; same, descending limb, 4). The band in gels 2 and 3 anodal to the γ-dimer position represents monomeric DNS-cad-labelled γ chains.

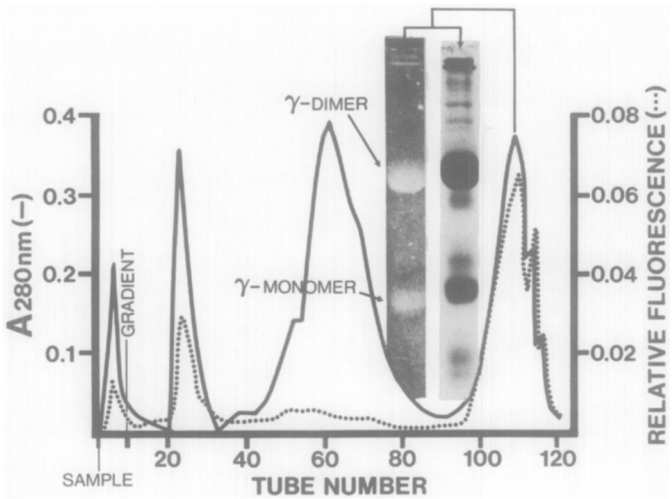

Fig. 4 DEAE-cellulose gradient elution chromatography in 8M urea of 23 mg DNS-cad labelled S-carboxylmethylated fibrin. See legend Fig.1 for general conditions. Fractions of 2.2 ± 0.1 ml were collected (total gradient volume, 225 ml); at the completion of the gradient, the column was flushed with limit buffer. An electrophoretic gel (Laemmli system) is shown of material from the γ chain peak photographed under UV light, left, and subsequently stained with Coomassie blue, right.